

**Molecular Resource Center of  
Excellence  
2018–2019 Annual Report to the  
Tennessee Higher Education  
Commission**

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## I. Executive Summary

The Molecular Resource Center (MRC) of Excellence is an essential and successful institutional core research service that was established in 1985. Dedicated to providing state-of-the-art molecular technologies, fostering collaborative and interdisciplinary research, and providing education relevant to advancing molecular technologies, the MRC supports the research mission and prominence of the State of Tennessee. The successful accomplishment of this mission has been a result of continuous funding from the Tennessee Higher Education Commission (THEC) for a significant portion of the MRC operations and continuous matching funds support from the University of Tennessee Health Science Center (UTHSC). MRC's services are essential for our investigators to maintain their research programs in spite of continuing constraints in federal funding levels. Over the past five fiscal periods, we have met our goals to provide high quality DNA sequencing, whole genome, or next-generation sequencing (NGS), microarray-based gene expression analysis and nucleic acid amplification by polymerase chain reaction (PCR) services to our core customers.

In FY19, the MRC generated slightly less total recovery than in FY18 (\$247,232 vs. \$264,401, respectively). In general, since FY15, core revenues have been declining or have been flat, year over year. As availability of extramural grant funding has continued to decrease or to plateau over this five-year period, the volume of samples processed has also decreased. In addition, as observed for FY18, the majority of investigators we serve now select commercial vendors for performing their whole-genome, next-generation sequencing (NGS) projects. These decisions are primarily based on cost sensitivity as there is reduced per

sample pricing widely available commercially. Commercial vendors operate high-throughput sequencers as opposed to the mid-throughput instruments housed in the MRC. Commercial vendors continue to compete with each other for academic business by offering “new customer” bulk order discount pricing, in addition to lower overall per sample prices. The type of MRC customers who have decided to send their NGS samples to commercial vendors do not typically need extensive experimental design consultation, or local expert advice on how to troubleshoot sample preparation, or assistance with downstream raw data analysis (they analyze their own data). In FY19, those customers with small to moderate sample sizes who rely on the MRC for the above expertise and services have continued to use the MRC to support their research program. In addition, we continue to serve customers who require rapid turnaround time; these investigators are time-sensitive rather than price-sensitive. To meet the budget constraints of our price-sensitive core users, we recently negotiated with Novogene to set up a blanket PO to offer their tier of discounted rates to all UTHSC investigators. This contract was approved in October of 2019. Novogene is one of the commercial vendors that absorbed our prior MRC NGS customers, including Dr. Rob Williams. Our goal will be to facilitate consultations with clients, advise on the pros and cons of staying in-house or choosing Novogene. Each research group will decide if it is better to use the MRC to take advantage of rapid turnaround time or to send out samples at a reduced per sample price, albeit with a 4-6 week turnaround time.

In FY19, relative to FY18, the revenues generated from NGS increased by ~16% (\$58,908 to \$69,511, respectively). The largest volume user of NGS services was Dr. Neil Hayes during this period. In FY19, 389 library preps and 52 Proton and Illumina sequencing runs were completed versus 139 library preps and 51 Proton

and Illumina sequencing runs completed in FY19, a 53.5% decrease in overall sample number. Note that more libraries can be sequenced on an individual “run” on a NextSeq instrument since higher capacity cartridges are available for this platform versus the Proton. For the first time since acquiring the Illumina NextSeq, more samples were processed on the NextSeq instrument than the Proton (41 Illumina runs vs. 10 Proton runs). In FY19, the number of “runs” was similar, but fewer library preps were requested, likely because the Hayes laboratory at UTHSC generates their own library preps prior to sequencing in the MRC. The MRC customers who no longer use the MRC for NGS services are concentrated in the Department of Genomics, Genetics & Informatics, including one of our prior top five core users, Dr. Rob Williams (GGI Chair). When Dr. Williams began sending the majority of his group’s whole-genome NGS projects to outside vendors, external customers he had referred to the MRC followed suit, including his collaborator, Dr. Klaus Schugart (Helmholtz Center for Infectious Diseases, who also has a joint appointment in the MIB department).

In terms of volume and/or recovery, the demand for Agilent Bioanalyzer services decreased from 312 processed samples in FY18 to 257 processed samples in FY19, leading to decreased revenues. Although recoveries from Affymetrix microarrays decreased by 43% (FY18: \$79,167 to FY19: \$44,818), arrays continue to be an important method for generating differential gene expression data for our customers, particularly those customers with constrained budgets that cannot afford to use NGS methods.

In FY19, revenues from Sanger sequencing significantly increased (FY18: \$17,530 to FY19: \$42,967). The demand for Qiagen QIAcube automated nucleic acid

preparation was stable from FY18 to FY19, whereas the demand for real-time PCR services and reagents for the Roche LC480 platform decreased by 16% as compared to FY18 (FY19: \$34,989 to FY18: \$41,723). Overall, FY19 ended with a net income of \$54,197, similar to FY18, which ended with a net income of \$31,338.

In FY19, MRC processed >19,500 service units on behalf of 111 total investigators (94 internal and 17 external users) across 18 departments and five Colleges at UTHSC, and seven unique external academic institutions (including University of Memphis, Christian Brothers University, University of Nebraska and Boston University) and one local commercial entity, Silver Bullet Lab, Bartlett, TN. In FY19, the MRC contributed to >240 unique peer-reviewed publications and review articles. The Director, Dr. Taylor, provided >80 consultations to core users and multiple letters of support for grant applications. Dr. Taylor also provided core facility tours to 14 prospective faculty members and to VIP campus visitors.

Due to continuing increased costs of personnel (routine salary increases) and service contracts projected for FY20, compounded by decreased demand for NGS and microarray services, we do not anticipate that any THEC funds will be used to purchase new major equipment (>\$10,000) in FY20; therefore, equipment needs will need to be subsidized directly by the UTHSC campus. Equipment needs will be re-evaluated in Q3 and Q4 of FY20 to determine if there are sufficient funds to support equipment upgrades or purchases. It is predicted that MRC total recoveries will remain flat in FY20 relative to FY19.

In FY20, there remains interest in purchasing the 10X Genomics Q3 instrument for single cell sequencing applications. The MRC and Dr. Seagroves will continue to work closely with Dr. Steven Goodman, the Vice Chancellor for Research, to manage investments in emerging molecular technologies. A primary mission of the Executive Director, the Associate Vice Chancellor for Research—Core Labs, and the Office of Research senior administration going forward will also be to ensure that matching funds are available to supplement the portion of the MRC operating budget provided by the THEC.

The MRC continues to enhance its services with tools and procedures to increase productivity, as well as increasing the user's understanding of MRC services, as key components in investigator-initiated research. The MRC's investment in state-of-the-art analytical tools and procedures provides researchers with a competitive advantage for new and renewal grant applications, providing a clear return on investment (ROI). In order for all UTHSC campus investigators to take advantage of the THEC's investment in the MRC, we also support educational programs about current and emerging technologies. In FY19, the MRC leveraged the support of the State of Tennessee to provide services to the UTHSC research community with value in excess of \$3.0M, which is conservatively estimated based upon "street-value" of services provided, particularly for our microarray profiling services. The UTHSC MRC is one of the few remaining academic core centers in the country that provides microarray services. Microarrays continue to be a high-throughput alternative to NGS, particularly for researchers who are primarily interested in differential gene expression of moderate to highly expressed mRNAs, but not low abundance mRNAs, miRNAs or differential isoforms of the same gene.

In 2015, MRC expanded its services to include a new, “in-house” research bioinformatics core for molecular informatics (Molecular Bioinformatics, mBIO) that was initially formally housed under the MRC umbrella. The core is directed by Dr. Daniel Johnson, who was recruited to campus in May 2015. The mBIO core successfully addressed a prior unmet need on our campus, which is the analysis of large-scale data generated in the MRC from microarray and NGS-based profiling studies at a subsidized rate. The mBIO unit provides pre-experimental design consultation (at no charge) and post-experiment data analysis (at a subsidized hourly rate) to all UTHSC campus investigators, including statistical analysis support after the raw data are generated by the MRC. In February 2016, Dr. Goodman accepted the Vice Chancellor for Research Cabinet’s recommendation to reassign the mBIO core a separate institutional core facility that is distinct from the MRC. Since the mBIO core is directly managed by the Office of Research, its activities are not included in the MRC THEC report. However, because the mBIO core resides in the same physical footprint as the MRC, this allows seamless generation and analysis of NGS and microarray data. The mBIO and MRC Core Directors and staff work closely together to assist investigators in execution of their research programs by jointly participating in all initial experimental design consultations. When the mBIO core advertises its services through Spring and Fall intensive workshops, the demand for NGS and array services at the MRC increases concordantly.



## **II. Mission Statement**

The mission of the Molecular Resource Center of Excellence (MRC) at The University of Tennessee Health Science Center is to extend and to enhance the molecular and cellular research capabilities of the research community by providing access to the latest technologies for exploring the molecular basis of health and disease. To provide these technologies, the MRC invests in equipment, trained personnel, and education. In addition, because the MRC is allied to all basic science and clinical science research units throughout the state, a component of this mission is to serve as a nucleus for collaboration and interdisciplinary research within the State of Tennessee.

*Tools for Success.* A fundamental objective of the MRC is to provide the research community with molecular tools that are essential for successful research. This is a dynamic process of assessment and anticipation to provide the intellectual and physical infrastructure that is responsive to the needs of the research community. With this objective in mind, the MRC has developed a staff of a faculty-level Director, three research staff and one administrative/support staff member to help researchers obtain the greatest value from their interactions with the MRC. The MRC and its staff have invested significantly in tools and facilities to provide high throughput sequencing of nucleic acids, large-scale analyses of the genes expressed in healthy and diseased organisms, ranging from bacteria to human, analyses of the genome of organisms, computational tools to analyze complex data, and the use of unique models of human disease. This intellectual and physical infrastructure that has developed around the MRC allows researchers to be nimble in their research and to move forward at a pace that sets a benchmark for competitiveness in biomedical research. Moreover, the MRC is listed as an

essential core facility and institutional resource on the majority of grant proposals submitted by our users, including investigators with multiple R01 or similar extramural awards, such as Dr. Philip (Dave) Rogers, Department of Clinical Pharmacy, Dr. Gadiparthi Rao, Department of Physiology and Dr. Subhash Chauhan, Department of Pharmaceutical Sciences. Therefore, the MRC is an essential component of research infrastructure on the UTHSC campus. The MRC also serves several external principal investigators through contracts negotiated with external universities located in the Memphis area (Rhodes College and Christian Brothers University), the Mid-South region, across the United States and around the world.

*Expert Staff.* The focus of MRC activities is to stimulate the development and dissemination of new methodologies and procedures as they become available to the scientific community. To stay abreast of current and emerging technologies, the MRC staff actively participates in professional development activities. These include formal activities such as scientific or professional development meetings and seminars. Other development activities are unstructured, such as reading journals and one-on-one or small group interaction with other scientific and technical personnel. In addition, the MRC staff devotes considerable time to developing novel protocols, scaling established protocols to minimize reagent costs and refining techniques, analyses, and processes.

The Associate Vice Chancellor for Research—Core Labs, Dr. Tiffany Seagroves, is also a user of MRC resources. She has been consistently extramurally funded as a cancer researcher since 2005. In addition to “wet bench” research and laboratory management skills, she brings additional management and operating

expertise to the institutional core facilities through her M.B.A. training. Dr. Seagroves has attended the Association for Biomolecular Research Facilities (ABRF) annual national meeting since 2015 to remain informed about core management best practices and strategies, including federal fiscal policies related to core operations. In 2018, Dr. Seagroves was elected to the executive board of the ABRF's regional chapter for core facilities, the Southeastern Association of Shared Resources (SEASR), and she will serve a four-year term on the board. Dr. Seagroves currently serves as the chapter President-Elect and she will help plan all annual SEASR meetings for the SE region related to shared resources. In 2020-2021, she will serve as SEASR President and will be primarily responsible for planning the 2021 annual SEASR meeting. She will encourage core staff from the MRC, throughout UTHSC and the State of Tennessee to attend the upcoming SEASR annual meeting in June 2020 at the Emory Conference Center in Atlanta, GA in order to advertise the MRC, and other UTHSC institutional core services, and to provide career development opportunities to our core personnel. The mBIO and MRC cores were advertised at the 2019 annual SEASR meeting, and UTHSC was a gold level institutional sponsor of the meeting.

Dr. Seagroves also serves on both the Vice Chancellor for Research, Research Cabinet and the Operational Strategic Plan for Research (OSPR) committees. The Research Cabinet advises the VCR on all activities relevant to research and the OSPR committee is charged with developing and executing a 5-year plan to strengthen core facilities and other research infrastructure units on the UTHSC campus, with the goal of aligning projected research needs and priorities with core capabilities. The OSPR was developed over FY16 and was published in September 2016. We are currently in the third year of execution of the strategic

plan. Supporting core facilities on campus is a major initiative highlighted in this strategic plan. As the largest molecular core facility on campus, the MRC will be an integral stakeholder in executing this strategic plan.

*Education and Training.* Another important part of the MRC mission is to provide educational opportunities. In an academic institution at the cutting edge of biomedical research, investigators must stay abreast of the technologies available; the MRC serves to focus this continuing education on molecular and cellular technologies. Fulfillment of the education mission has three goals: 1) develop a skilled workforce in the biomedical sciences, 2) keep investigators abreast of current and advancing technologies, and 3) allow investigators to discover new avenues to propel their research programs that ultimately shape the MRC's investment in new technologies.

Workforce development is vital to the future of biomedical sciences research and industry in the State of Tennessee. For many of its functions, the MRC serves as a teaching laboratory for undergraduate and graduate students, technical staff, and investigators. The MRC staff instructs these users one-on-one at the benchtop and then provides guidance as new core users develop their skills. Because the MRC continually monitors advancing technologies to anticipate needs of investigators, it is in the best position to provide educational programs to investigators about the use of current technologies and technologies that are on the horizon. Each year, the MRC hosts seminars and workshops on topics designed to enhance progress in the UTHSC community by raising awareness of new molecular tools that are available. The investigators served by the MRC are always looking for improved strategies to answer their scientific questions.

*Assessing Needs and Satisfaction.* Through frequent dialog with the investigators we serve, and meetings of the core internal advisory board (IAB), the MRC also investigates specific investments in, or adaptation of, technologies to best meet investigator needs. Needs are also brought to the MRC's attention for consideration through individual principal investigators, the IAB and the Office of Research. Needs that cannot be currently met by MRC are referred to the Associate VCR, the Senior Associate VCR and/or the VCR for further consideration. Campus survey tools are in place to inquire about research needs on campus and to gauge the overall satisfaction with individual core services.

*Collaboration.* The MRC is an integrative unit that is egalitarian in its delivery of services across all units within the Tennessee research community. In this role, the MRC fosters collaboration and the development of interdisciplinary research. For example, the MRC sponsors seminars and workshops in genomics, bioinformatics, microarray technologies and other molecular technologies. In addition, the MRC has established an active participation in seminars/webinars offered by the ABRF or SEASR. In collaboration with the Center for Integrative and Translational Genomics (CITG), the MRC has underwritten and developed numerous collaborations that use state-of-the-art molecular technologies. Likewise, the MRC is playing a significant role in the development of large-scale collaborations to understand complex gene interactions through mouse models of human disease. These highly visible efforts have placed The University of Tennessee in a position whereby researchers may take fullest advantage of funding and research opportunities.

### III. Center of Excellence Overview

a. **Center Name:** Molecular Resource Center of Excellence

b. **Center Location:**

- i. Administrative Address –  
University of Tennessee Health Science Center  
Translational Science Research Building (TSRB)  
71 S. Manassas St., Room 110  
Memphis, TN 38163

**Web address-**

<https://www.uthsc.edu/research/institutional-cores/mrc/index.php>

- ii. Laboratory Address –  
Molecular Resource Center  
Translational Science Research Building (TSRB)  
71 S. Manassas St., Room 110  
Memphis, TN 38163

c. **Name and Title of Person Responsible for Administering Center:**

Tiffany N. Seagroves, Ph.D.  
Executive Director

Associate Vice Chancellor for Research—Core  
Labs  
Professor of Pathology  
College of Medicine  
University of Tennessee Health Science Center

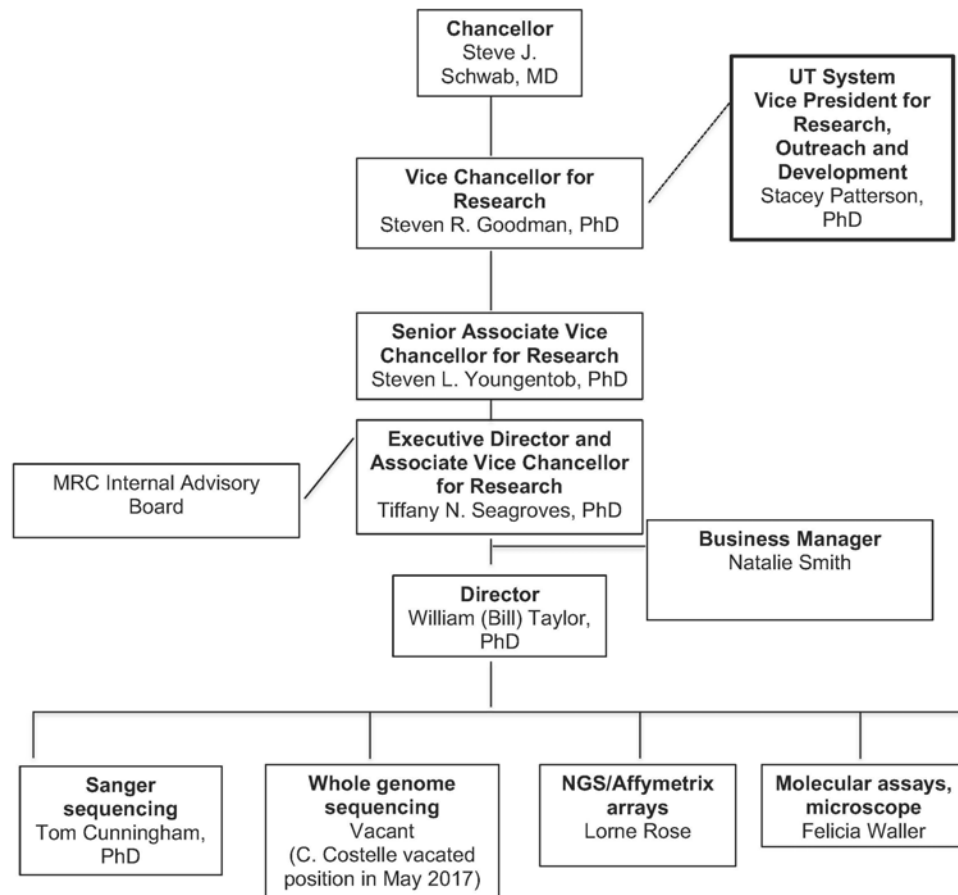
d. **Unit Which Houses Center:**

Office of Research  
Vice Chancellor for Research  
University of Tennessee Health Science Center

Steven R. Goodman, Ph.D.  
Vice Chancellor for Research

Steven Y. Youngentob, Ph.D.  
Senior Associate Vice Chancellor for Research

*Organizational Structure.* The Molecular Resource Center (MRC) is located within the organizational structure of the University of Tennessee system under the Office of Research, which is directed by the Vice Chancellor for Research. The center itself is under the supervision of the Executive Director, Dr. Tiffany Seagroves, and the Core Director, Dr. Bill Taylor. Dr. Seagroves is a professor in the Department of Pathology in the College of Medicine. As Executive Director of the MRC, Dr. Seagroves reports directly to the Senior Associate Vice Chancellor for Research, Dr. Steven Youngentob, who reports to the Vice Chancellor for Research, Dr. Steven Goodman. Dr. Goodman reports directly to the Chancellor of the UTHSC, Dr. Steve J. Schwab.



The day-to-day operations of the Center are supervised by the Director of the MRC, Dr. William (Bill) Taylor, a full-time faculty-level staff member of the Molecular Resource Center of Excellence, and Dr. Tom Cunningham, Instructor and Associate Director of the MRC. In FY16, a new Internal Advisory Board (IAB), comprised of faculty members representing campus departments that use the services of the MRC, was appointed by Dr. Goodman. The IAB advises the VCR how MRC funds should be used and makes recommendations related to new equipment requests for the MRC, although final decisions are made by the Executive Director in consultation with the Vice Chancellor for Research.

**MRC Internal Advisory Board, FY19**

<b>PI:</b>	<b>Department Affiliation:</b>
Ramesh Narayanan, Ph.D.	Medicine
Lawrence Reiter, Ph.D.	Neurology
Lu Lu, Ph.D.	Genetics, Genomics and Bioinformatics
P. David Rogers, Pharm.D., Ph.D.	Clinical Pharmacy ( <i>IAB CHAIR</i> )



#### **IV. Year End Review FY19**

Receivables for core services decreased slightly in FY19 as compared to FY18. In FY19, the MRC processed >19,500 independent samples/supply requests (not including use of equipment available in the MRC at no charge). During this fiscal period, the MRC served 111 total investigators, of which 94 investigators were internal to UTHSC (84.7% of all investigators), representing the core of our currently extramurally funded investigators, and 17 were external users, representing seven academic institutions and one commercial vendor. The large number of faculty served is reflective of our continuing commitment in the MRC to providing excellence in molecular analyses, while promoting significant advances in our services related to nucleic acid sequencing, educational programs, and fostering collaboration and communication.

Over the past several years, the MRC committed itself to the rapidly evolving technologies of high-throughput, massively parallel nucleic acid sequencing (next-generation sequencing (NGS), also known as whole genome sequencing. To recapitulate the impact of these technologies, an individual investigator can now sequence nucleic acids on a scale far exceeding that of hundreds of investigators in the human genome project of the previous decade, and in a time frame of minutes and hours instead of weeks and months. Applying these technologies allow the rapid analysis of genomic DNA, expressed messenger RNA, active regions of chromosomes, regions of DNA coding for proteins, modification of DNA, and other, constantly evolving analyses.

Our first ABI Proton NGS instrument was installed and became functional in FY14. In FY15, based on our status as a beta testing site for the next generation PII Proton chip, Thermo/Life Technologies provided a second ABI Proton instrument on permanent “loan” to the MRC through FY18. Currently, the MRC offers two tiers of analysis on the Life Technologies platform, the PGM and the Proton (PI) chips, depending on the complexity of the proposed sequencing project. In FY17, an Illumina NextSeq 500 NGS instrument was purchased by the UTHSC VC for Research for the MRC. The Illumina NextSeq500 instrument was installed in July 2016, which added a platform option for NGS. Adoption of use of the Illumina NextSeq 500 platform has been slower than anticipated; however, use increased in FY19 relative to FY18 due to increased demand by investigators seeking rapid turnaround time on NGS projects. Effective FY17, the pricing for whole genome sequencing services was set to be platform-agnostic, allowing users to choose the platform based on technology preferences or their throughput needs, rather than pricing. The NextSeq500 high-throughput chip has the capacity to sequence up to 230 million base pairs, whereas the typical output of the Proton P1 chip is 80-90 million reads, allowing more samples to be profiled simultaneously on a single run using the Illumina NextSeq cartridge.

To improve the efficiency and consistency of many operations, the MRC is heavily invested in robotic technologies. The most significant labor-intensive step in NGS lies in the bar-coded library preparation. In December 2016, we installed the Hamilton Starlet robot dedicated to NGS library preparation, which had become a bottleneck to sample processing, and, therefore, to rapidly invoicing processed NGS samples. Instead of producing 16 libraries per week using all manual labor liquid handling, the robot is capable of generating >100 libraries per week. As of

FY18, the Starlet is fully programmed to prepare most NGS libraries on a large-scale, decreasing manual sample manipulation.

The expansion of NGS capabilities in the MRC, along with continued increased interest by investigators, has allowed the MRC to enhance its service delivery components. To educate potential users regarding use of the technology, the MRC continues to sponsor seminars to provide users with a starting point for experimental design. Dr. William Taylor, Director of the MRC and Dr. Daniel Johnson, Director of the Molecular Bioinformatics (mBIO) core are also critical assets to assist investigators with one-on-one planning of experimental design and subsequent interpretation of data. Consultation for project design and a general plan for statistical methods to be used for data analysis are provided at no charge to all core customers.

With the expansion of the Molecular Bioinformatics (mBIO) core, an independent institutional core facility located in the same physical footprint at the MRC, we have been able to reduce MRC's investment in some pre-packaged NGS analysis software tools, including the Partek platform, which previously allowed end-users to evaluate the quality of their NGS runs. The mBIO core also supports a local installation of the GALAXY tools for NGS data analysis, allowing PIs to use standardized workflows to analyze their data without investing in customized bioinformatics workflows. Therefore, we support a breadth of investigator needs and capabilities in raw data analysis.

The gap between price points and ease of data analysis between microarray technology and NGS is significant, so the MRC continues to provide array

platforms to allow the quantitation of gene expression at or near single transcript levels using microarrays (Affymetrix or Illumina iScan arrays). Of note, unlike microarrays, there are no standardized methods for NGS “runs” or downstream data analysis workflows for research on deidentified, exempt human clinical samples, which is another advantage of using arrays. The MRC is one of a few academic core facilities in the US that continues to offer microarray services for gene expression profiling, so it is possible that use of our core by external users may increase over time. Multiplexed PCR reactions (96 samples x 96 primer sets), and digital PCR are also expected to be increasingly popular to validate array and NGS experiments, using the Fluidigm Biomark instrument.

Conventional capillary (Sanger) sequencing of DNA continues to account for a large proportion of the total number of samples processed in MRC. This service, directed by Dr. Tom Cunningham, processed 8,684 samples in FY16, 7,979 samples in FY17, 7,394 samples in FY18, and 9,272 samples in FY19. In FY15, we also began offering direct shipping to an external sequencing vendor, GeneWiz. In FY19, 1,877 samples were sent to GeneWiz for Sanger sequencing, a significant increase over the FY18 volume of 417 samples. Competitive pricing for GeneWiz was fixed per the initial contract until December of 2017. In FY18, a three-year pricing contract with GeneWiz was renegotiated and signed at FY17 price levels, so that we could retain these services through 2021. If Sanger sequencing is ever sunsetted at MRC, then all samples could be easily transferred to the GeneWiz dropbox at the MRC for external processing at these negotiated prices.

In general, Sanger sequencing has become a commodity, and several other academic genomics cores nationwide have sunsetted this service, investing their personnel resources in NGS or other technically challenging molecular services. Throughout FY18-FY19, with the MRC unable to recruit for the advertised open position for NGS services that was vacated by Ms. Costelle, Dr. Cunningham has assisted with microarrays, allowing Lorne Rose to learn NGS protocols. If the MRC open position is not filled, it will be important to retain Dr. Cunningham to assist with MRC operations. Therefore, the decision regarding sunseting Sanger sequencing is deferred until FY21.

Although sample numbers for microarray processing services decreased from FY18 to FY19 (1,056 to 264 samples), microarrays remain an important discovery tool for our researchers. The microarray services, managed by Mr. Lorne Rose, primarily use the Affymetrix platform. The iScan (Illumina) microarray system that is primarily used for genotyping, identification of SNPs and ethnicity profiling was installed in December 2015. Although there was no use of the iScan platform in FY16, 32 samples were processed in FY17, and 80 samples were processed in FY18 and 32 samples were processed in FY19 (processing fee only, since the end user purchased his own reagents directly from the vendor). From FY16-FY19, only the laboratory of Dr. Dave Shibata (Chair, Surgery) has used the iScan instrument. Therefore, in Q1 of FY19, the instrument was moved to his personal laboratory space. The instrument is not currently maintained under a maintenance service contract due to low volume usage.

The number of real-time polymerase chain reaction (qPCR) samples processed decreased in FY18 (from 541 runs in FY18 to 361 runs in FY19). Several PIs now

house their own benchtop qPCR instruments since the prices for these instruments have dropped over the past 5 years. The PCR operations, overseen by Ms. Felicia Waller, continue to have solid bookings, and several investigators also buy their PCR reagents (master mixes and PCR plates) directly from the MRC. For example, \$31,299 in recoveries were generated in FY19 from the resale of qPCR reagents, whereas \$3,690 in recoveries were generated for qPCR instrument use (plate run service fees).

Although there were 16 Fluidigm Biomark microfluidics real-time PCR experiments supported in FY19, this service generated no revenues since all users provided their own reagents and operated the instrument on their own. Several investigators are interested in single cell technologies and development of a single cell analysis institutional core is proposed in our new strategic plan for research. The Biomark system allows the robotic mixing of up to 96 sample with up to 96 primer pairs (i.e., 96 analyses of 96 samples), for a total of 9,216 analyses in a single run lasting just a couple of hours. We anticipate increased usage of the Biomark real-time PCR system for validation of “hits” identified from NGS of sequencing single cells processed on the Fluidigm C1 single cell analysis instrument currently housed in the GGI Department.

The business administration of MRC is led by Ms. Natalie Smith; she is responsible for a host of functions, ranging from contracts and invoicing core users, to seminar, workshop, and conference scheduling. To help facilitate sample tracking, investigator billing, and institutional research, in FY17 a major effort to move the accounting system to a more robust and easily supported software system was spearheaded by Dr. Seagroves. In March 2017, the Office of Research entered

into an agreement with iLab Solutions (Agilent) to develop a web-based core laboratory management system that will facilitate requesting orders by PIs, invoicing of PIs by the institutional cores' business managers and reporting on core usage by the Associate Vice Chancellor for Research. iLab users use single sign on authentication (NetID) for all active UTHSC investigators to provide access to services at all institutional core facilities. The MRC went live with iLab in January 2018. Since then, investigators have been able to request services, to track their project workflows, or to view their invoices within iLab. In addition, there is now financial integration of iLab with IRIS, such that only active funds may be selected by PIs to request core services. FY19 was the first full fiscal year that usage and revenues data could be easily analyzed for the MRC using the iLab software interface. Data may be classified and reported by individual service requester, the PI's lab group, department, month or individual service line, for example.

Beginning in July 2019, researcher funds will be automatically debited via IRIS through iLab after internal UTHSC and UTK approvals. This feature will mean that when the MRC business manager finalizes monthly invoices, funds will be debited immediately from a PIs designated account without the need to process internal transfer vouchers. Overall, adoption of iLab is expected to dramatically streamline the management of the MRC and to save hundreds of hours of labor per year related to processing invoices or to tracking down PIs to update incorrect information written on paper service request forms. Moreover, as of January 2018, after the iLab "go-live" date for MRC, paper service request forms were eliminated and all core service requests, all email communications sent via iLab and all invoices are permanently recorded in the iLab database, which is hosted in the cloud by Agilent.

In summary, the MRC touches many investigators at many institutions. The robust performance of MRC operations reflects a growth in the incorporation of new technologies and a steady user base in the State of Tennessee, the U.S., and abroad. This, in turn is a function of the dedicated and skilled staff as detailed in the organizational chart (Section III. Center of Excellence Overview).



## **V. Goals and objectives, FY20 (2019-2020)**

The goals of the MRC in FY20 are to fulfill its mission to extend and enhance the molecular and cellular research capabilities of the Tennessee research community and to provide an education resource to that community. There are three objectives to meet these goals:

*Expand the educational component of the MRC.* With technological advances occurring daily, the MRC will continue to provide educational programs to researchers to keep them abreast of developments, both current and those anticipated in the near future. The MRC will continue to host vendor seminars on emerging technologies and inviting guest speakers with expertise in genomics and molecular biology to the campus. The MRC will also attempt to assist community K-12 education with exposure to molecular research technologies and research. For example, there may be opportunities for interacting with the newly opened Crosstown High School charter school, which is located less than one mile from the UTHSC campus. The MRC will continue to host interns from the Southwest Tennessee Community College Biotechnology Technician program for their 14-week practical experience. SWTCC is located directly across the street from the MRC. Together, these educational initiatives enhance the competitiveness of research associated with the MRC and assist in workforce development, leveraging state support to strengthen biomedical research and research training. Since FY16, MRC has participated in “Hot Topics in Research” seminars organized by the Office of Research; one of these seminars highlighted features of the Illumina iScan and NextSeq500 instruments, and the second seminar highlighted the exciting preliminary NGS data recently generated in the MRC by the

laboratories of two MRC IAB members, Dr. Lawrence Reiter and Dr. Dave Rogers. The MRC also participated in several faculty recruitment visits, visits to UTHSC by external Office of Research administrators to share ideas about core management, hosted graduate student candidates, and celebrated postdoc week with UTHSC postdocs by offering a tour of MRC facilities.

*Expand awareness of MRC services to potential users throughout the State of Tennessee.* Although the majority of MRC users are located on or in close proximity to the UTHSC campus in Memphis, the MRC has been successful in “spreading the word” about the resources available to other researchers in the State. The MRC will continue to assist investigators throughout the State with their projects and project development and will attempt to reach additional investigators throughout the State through direct contact or informational programs. Dr. Seagroves and Dr. Jonathan Phipps, Director of the Core Facilities Program on the UT-Knoxville campus, met at the 2016 national ABRF meeting and they interact throughout the year to share ideas about core operations and to share information about the availability of core services available on our respective campuses. The overall goal of these types of contacts will be to increase awareness of cost-effective services provided by the MRC at UTHSC in Memphis, TN.

*Streamline new core management software use and improve core operations through enhanced metrics analysis.* As discussed, the MRC onboarded the iLab web-based system as of January 2018 at the MRC to manage core facility service requests and to track requests for services and to invoice core users. This required a large investment of core personnel time and effort to custom build their iLab

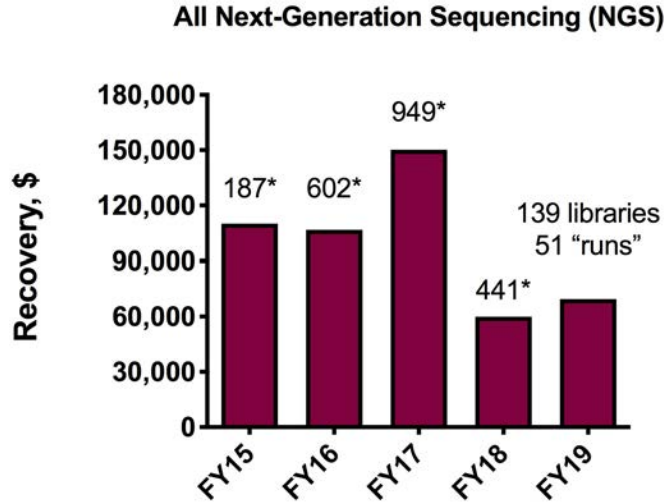
page, to create custom workflows to request services and to ensure that pricing was correctly entered into the database.

As predicted, the use of core management software has alleviated some of the issues we previously encountered in managing core records, which were maintained predominantly on paper from 1985-2017. For example, analysis of service use was very easy using the iLab interface for services provided from January to June 2018, however, it was more difficult to accurately pull the same information from the SlimPrim database for Q1-Q2 of FY18. SlimPrim has since been taken offline and all data contained within are now warehoused by the Office of Research Electronic Research Administration (ERA) office by Mr. Steve Wills. Going forward, all data regarding MRC services and invoicing will be exclusively housed in the iLab database, which has very powerful built-in data analysis programs and graphing capabilities. In terms of invoicing core users, using iLab has also alleviated issues with mis-spelling of PI names, use of incorrect internal account numbers, use of outdated internal account numbers and has allowed PIs to “join” each other’s laboratories to share funds assigned to a single PI. Communications about project workflow are also conducted via iLab and all messages are permanently stored, which will be useful in case of a future dispute about projects completed in the MRC. iLab also supports creation of custom quotes that can be accepted or rejected by either the PI or the core facility. A one-page brochure has been developed for each institutional core, including the MRC, and can be downloaded from the UTHSC Institutional Cores website. These brochures have been collated into a full-color, printed booklet highlighting the Institutional Core facilities. This document is updated on a yearly basis and has been shared with internal PIs at several faculty events over the course of 2018-

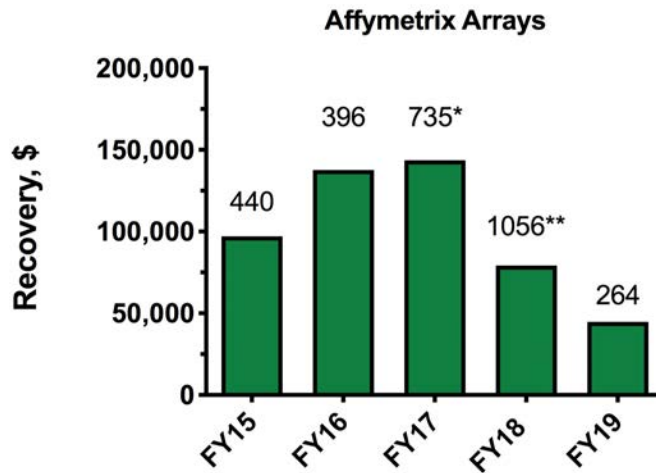
2019 and with postdocs during postdoc appreciation week and with individual visiting faculty candidates. The MRC website was also re-designed and streamlined in FY17. The MRC annual reports will be posted going forward to the MRC website (<https://uthsc.edu/research/institutional-cores/mrc/index.php>).

## VI. Sample Volume and Financial Summaries

### Review of FY19 Sample Volumes and Core Recoveries by Application:

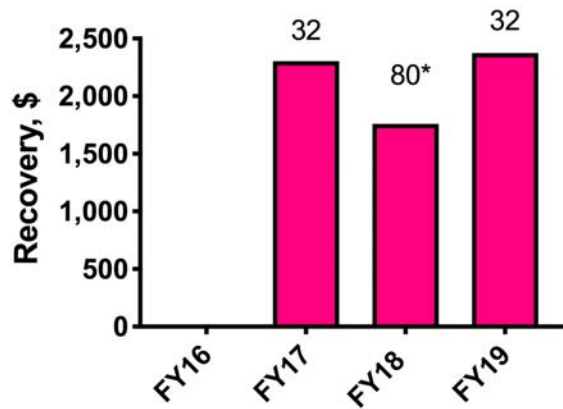


**Figure 1:** Five-year summary of recovery (\$) and total NGS usage (sequencing run + NGS library prep sample numbers= total number of services, as listed above each bar) for the ABI Proton, ABI PGM or the Illumina NextSeq500 NGS platforms. The recoveries include all NGS platforms and use data includes the total of library preps plus the number of sequencing “runs”. In FY17, library prep accounted for the majority of sample volume, whereas in FY19, fewer total libraries were prepared, but more high-throughput cartridge “runs” on the Illumina NextSeq were requested.



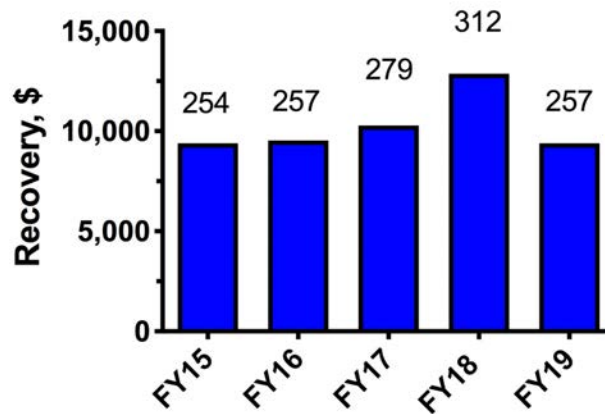
**Figure 2:** Five-year recovery (\$) and usage (number of arrays processed is shown above each bar) for all Affymetrix microarray services. \*In FY17 100 individual arrays were purchased, but were not yet processed, leading to apparent increases in sample number. In FY18, the arrays purchased in bulk in FY17 were processed, leading to high service volume, but lower overall revenues from array services.

### Illumina iScan Arrays (new in FY16)

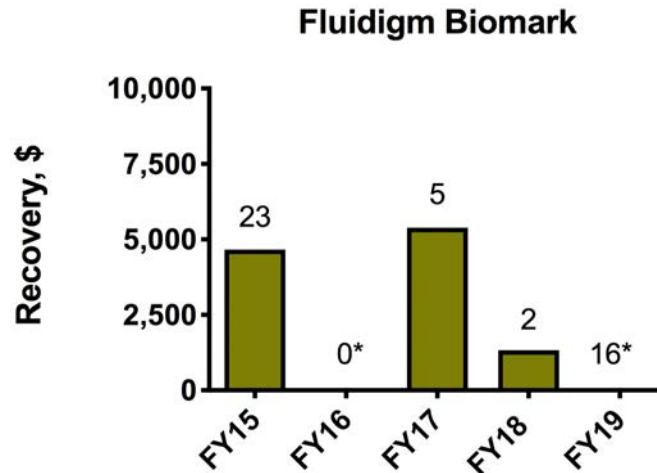


**Figure 3:** Five-year recovery (\$) and usage (number of samples processed on iScan arrays is shown above each bar) for all Illumina iScan microarray services. In FY17, arrays and reagents were purchased from the MRC, but in FY18, the single investigator who has used the iScan purchased reagents directly from Illumina, and the investigator paid just the array processing fees to the MRC.

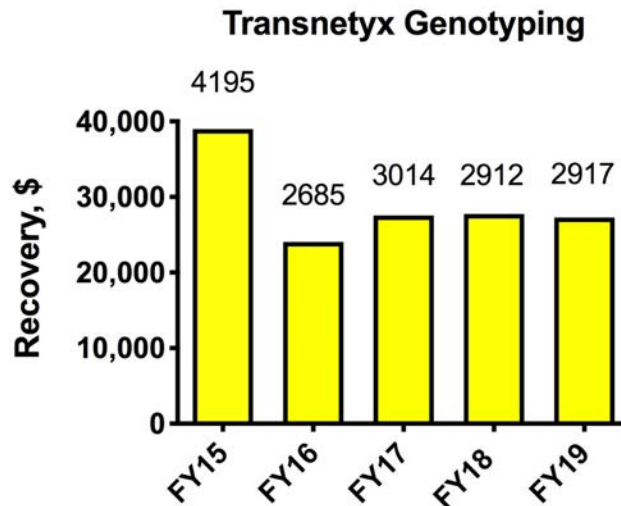
### Agilent Bioanalyzer



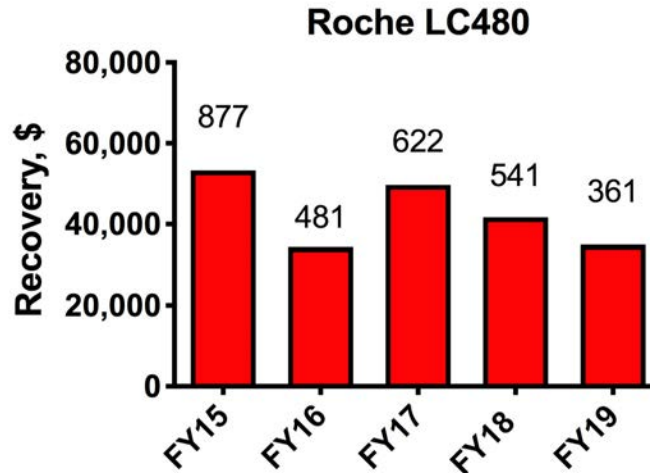
**Figure 4:** Five-year recovery (\$) and usage (number of samples processed is shown above each bar) for the Agilent Bioanalyzer assays, which quantitate DNA and RNA and/or evaluate RNA integrity. These numbers do not include any Bioanalyzer runs that were part of our standard quality control (QC) assessment for NGS runs. Customers who use outside NGS vendors often use the Bioanalyzer services before sending out their samples for library preparation/sequencing runs.



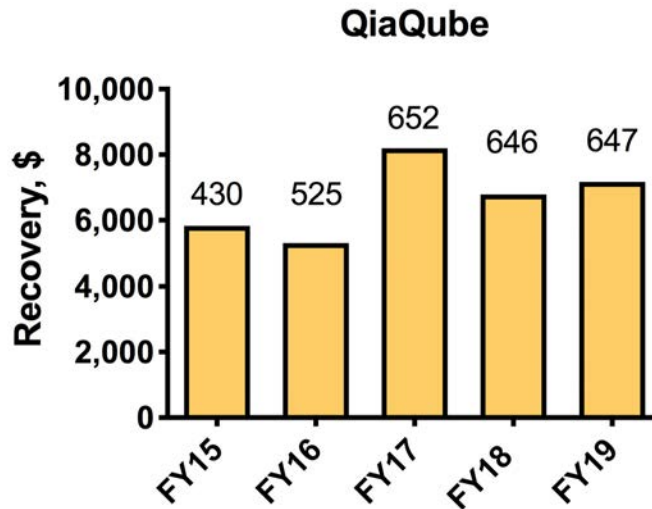
**Figure 5:** Five-year recovery (\$) and usage (number of projects processed is shown above each bar) for the Fluidigm Biomark multiplex PCR equipment. As our investigators expand into single cell capture and NGS profiling of single cells, or as they validate microarray/NGS gene expression data, this instrument is likely to be more heavily used for validation, as it is optimal for low input samples and moderate throughput gene expression analyses.



**Figure 6:** Five-year recovery (\$) and usage (the number of samples processed is shown above each bar) for the Transnetyx genotyping service. Usage has remained steady since FY16.

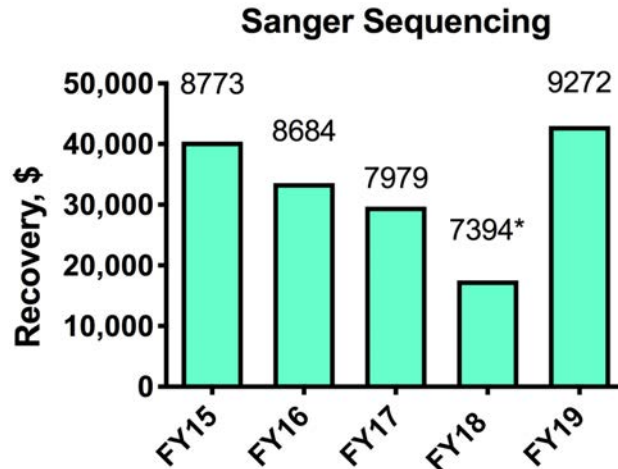


**Figure 7:** Five-year recovery (\$) for all qPCR reagents and Roche LC480 real-time PCR plate “runs” (the number of instrument runs shown above each bar). As more investigators purchase their own qPCR instruments, and as users move towards the higher density 384-well format, the total number of plates processed, and the amount of reagents purchased has decreased from FY15 levels. In FY17, the price per plate run was increased by 100% (from \$5/run to \$10/run), leading to a significant increase in revenue relative to FY16. Instrument use has declined by almost 50% since FY17 as more investigators purchase their own benchtop qPCR instruments.

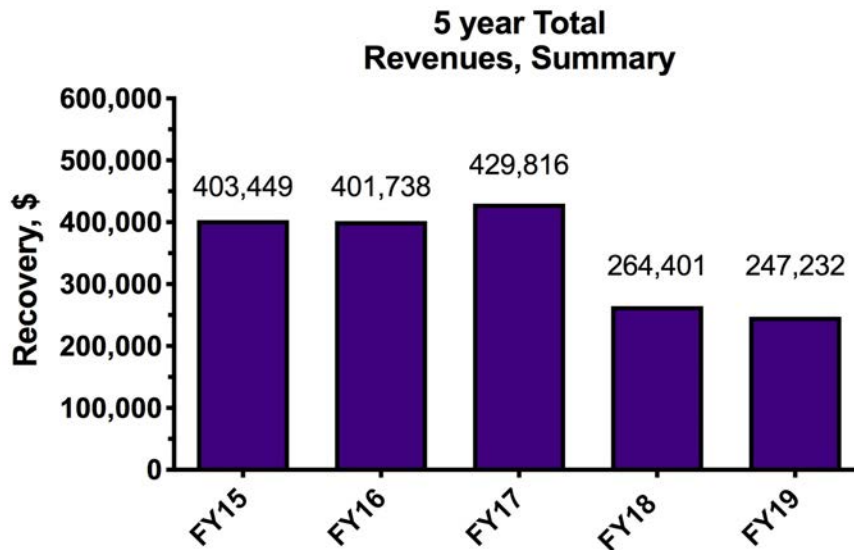


**Figure 8:** Five-year recovery (\$) and usage (number of samples processed is shown above each bar) for the automated preparation of nucleic acids from cells, fluids or pulverized tissues using the Qiagen QiaQube instrument. For enhanced quality control prior to NGS library preparation for RNA-seq, investigators are encouraged by MRC staff to utilize the QiaQube for their preparations of DNase-treated RNA, which will be free of contaminants typically observed with other standard laboratory RNA extraction methods.





**Figure 9:** Five-year recovery (\$) and usage (number of samples processed is shown above each bar) for Sanger sequencing. From FY15-FY18, sample number was slowly decreasing; however, sample volume increased in FY19. There is competition due to the option of external vendor provider (MRC is a also dropbox for GeneWiz), and due to the increasing commerical availability of fully-sequenced “ready to use” clones and artificial gene synthesis services, which reduce the need to sequence DNA to verify the correct orientation or sequence identity.



**Figure 10:** Total recovery (\$) for the MRC since FY15. Overall, revenues have were stable from FY15-FY17. Recoveries significantly decreased in FY18 over prior FY periods as more customers selected commercial vendors for NGS services while the use of microarrays also declined. NGS vendors are able to compete with academic cores using aggressive discounted pricing per sample because they use of higher throughput sequencing platforms than the MRC (economy of scale).

## VII. MRC Assisted Investigators FY19, July 2018-June 2019

<b>Principal Investigator:</b>	<b>Department:</b>	<b>College:</b>
Armstrong, William E	Anatomy & Neurobiology	Medicine
Chizhikov, Viktor V.	Anatomy & Neurobiology	
Fletcher, Max	Anatomy & Neurobiology	
Foehring, Robert	Anatomy & Neurobiology	
Hamre, Kristin M.	Anatomy & Neurobiology	
Heck, Detlef H.	Anatomy & Neurobiology	
Reiner, Anton	Anatomy & Neurobiology	
Youngentob, Steven L.	Anatomy & Neurobiology	
Zhang, Yanhui	Bioscience Research	Dentistry
Fortwendel, Jarrod	Clinical Pharmacy	Pharmacy
Palmer, Glen	Clinical Pharmacy	
Peters, Brian	Clinical Pharmacy	
Rogers, P. David	Clinical Pharmacy	
Freeman, Kevin	Genetics, Genomics and Informatics	Medicine
Jones, Byron	Genetics, Genomics and Informatics	
Lu Lu	Genetics, Genomics and Informatics	
Mulligan, Megan	Genetics, Genomics and Informatics	
Simpson, Claire	Genetics, Genomics and Informatics	
Starland-Davenport, Athena	Genetics, Genomics and Informatics	
Williams, Robert W.	Genetics, Genomics and Informatics	
Gerling, Ivan	Medicine	
Hayes, David Neil	Medicine	
Kang, Andrew	Medicine	
Makowski, Liza	Medicine	
Miranda-Carboni, Gustavo	Medicine	
Narayanan, Ramesh	Medicine	
Quarles, Leigh D.	Medicine	
Reed, Guy L.	Medicine	
Roslonicc, Edward	Medicine	
Cooper, Terry	Microbiology, Immunology and Biochemistry (MIB)	Medicine
Dale, James	MIB	
Fitzpatrick, Elizabeth	MIB	
Gomes-Solecki, Maria	MIB	
Guntaka, Ramareddy	MIB	
Hori, Roderick	MIB	
Jonsson, Colleen	MIB	
Kong, Ying	MIB	
Marion, Tony	MIB	
Radic, Marko	MIB	
Schughart, Klaus	MIB	

Khan, Mohammad	Neurology	Medicine
LeDoux, Mark S.	Neurology	
McDonald, Michael	Neurology	
Reiter, Lawrence	Neurology	
Tsao, Jack	Neurology	
Stanfill, Ansley Grimes	Nursing	Nursing
Gangaraju, Raja	Ophthalmology	Medicine
Jablonski, Monica	Ophthalmology	
Morales-Tirado, Vanessa	Ophthalmology	
Cho, Hongsik	Orthopaedic Surgery	Medicine
Miranda (Krum), Susan	Orthopaedic Surgery	
Larabee, Ronald	Pathology	Medicine
Pfeffer, Lawrence	Pathology	
Seagroves, Tiffany N.	Pathology	
Wu, Zhaohui	Pathology	
Astreinidis, Aristotelis	Pediatrics	Medicine
Bissler, John	Pediatrics	
Brown, Chester	Pediatrics	
Chen, Guoyun	Pediatrics	
Han, Joan	Pediatrics	
Mroczkowski, Henry	Pediatrics	
Pierre, Joseph F.	Pediatrics	
Purejav, Enkhsaikhan	Pediatrics	
Ryder, Alex	Pediatrics	
Samarasinghe, Amali	Pediatrics	
Towbin, Jeffrey A.	Pediatrics	
Zhang, Weiqiang	Pediatrics	
Chauhan, Subhash	Pharmaceutical Sciences	Pharmacy
Hevener, Kirk	Pharmaceutical Sciences	
Jiang, Jianxiang	Pharmaceutical Sciences	
Kurosu, Michio	Pharmaceutical Sciences	
Li, Wei	Pharmaceutical Sciences	
Petkov, Georgi	Pharmaceutical Sciences	
Yallapu, Murali	Pharmaceutical Sciences	
Bahouth, Suleiman	Pharmacology	Medicine
Bukiya, Anna	Pharmacology	
Chen, Hao	Pharmacology	
Dopico, Alex	Pharmacology	
Jee, Chang Hoon	Pharmacology	
Liao, Francesca-Fang	Pharmacology	
Malik, Kafait U.	Pharmacology	
Park, Ed	Pharmacology	
Raghow, Rajendra	Pharmacology	
Tavalin, Steve	Pharmacology	

Carson, James	Physical Therapy	Allied Health
Cordero-Morales, Julio	Physiology	Medicine
Jaggur, Jonathan	Physiology	
Mancarella, Salvatore	Physiology	
Rao, G.N.	Physiology	
Sun, Zhongjie	Physiology	
Tigyi, Gabor	Physiology	
Vasquez, Valeria	Physiology	
Mozhui, Khyobeni	Preventive Medicine	Medicine
Gosain, Ankush	Surgery	Medicine
Mas, Valeria	Surgery	

### **MRC-Assisted External Investigators**

#### **Principal Investigator:**

Abell, Amy  
 Baker, Daniel L.  
 Brown, Shawn  
 Ciobanu, Daniel  
 Fitzgerald, Jonathan  
 Frawley, Elaine  
 Gabai, Vladimir  
 Jackson-Hayes, Loretta  
 LeDoux, Mark  
 Mandel, Jennifer  
 Olgivie, Mary  
 Pandey, Udai  
 Sherman, Michael  
 Silver Bullet Biology  
 Sutter, Claire  
 Thompson-Jaeger, Sandra  
 Wheeler, Bayly

#### **Entity:**

University of Memphis, Memphis, TN  
 University of Memphis, Memphis, TN  
 University of Memphis, Memphis, TN  
 University of Nebraska, Lincoln, NE  
 Rhodes College, Memphis, TN  
 Rhodes College, Memphis, TN  
 Boston University, Boston, MA  
 Rhodes College, Memphis, TN  
 University of Memphis, Memphis, TN  
 University of Memphis, Memphis, TN  
 Christian Brothers University  
 University of Pittsburgh, Pittsburgh, PA  
 Vanderbilt University, Nashville, TN  
 Bartlett, TN  
 University of Memphis, Memphis, TN  
 Christian Brothers University, Memphis, TN  
 Rhodes College, Memphis, TN

## VIII. Publications Supported by the MRC in FY19 (published July 1, 2018 to June 30, 2019)

There were >240 unique publications supported by the MRC in FY19. Individual users of the MRC are bolded in the citations.

### **Astreinidis, Aristotelis**

Valianou M, Filippidou N, Johnson DL, Vogel P, Zhang EY, Liu X, Lu Y, Yu JJ, Bissler JJ, **Astrinidis A**. Rapalog resistance is associated with mesenchymal-type changes in Tsc2-null cells. *Sci Rep*. 2019 Feb 28;9(1):3015. doi: 10.1038/s41598-019-39418-5. PubMed PMID: 30816188; PubMed Central PMCID: PMC6395747.

### **Bahouth, Suleiman**

Nooh MM, Kale A, **Bahouth SW**. Involvement of PDZ-SAP97 interactions in regulating AQP2 translocation in response to vasopressin in LLC-PK(1) cells. *Am J Physiol Renal Physiol*. 2019 Aug 1;317(2):F375-F387. doi:10.1152/ajprenal.00228.2018. Epub 2019 May 29. PubMed PMID: 31141395.

Nooh MM, Mancarella S, **Bahouth SW**. Novel Paradigms Governing  $\beta(1)$ -Adrenergic Receptor Trafficking in Primary Adult Rat Cardiac Myocytes. *Mol Pharmacol*. 2018 Aug;94(2):862-875. doi: 10.1124/mol.118.112045. Epub 2018 May 30. PubMed PMID: 29848777; PubMed Central PMCID: PMC6022806.

### **Bissler, John**

Valianou M, Filippidou N, Johnson DL, Vogel P, Zhang EY, Liu X, Lu Y, Yu JJ, **Bissler JJ**, Astrinidis A. Rapalog resistance is associated with mesenchymal-type changes in Tsc2-null cells. *Sci Rep*. 2019 Feb 28;9(1):3015. doi:10.1038/s41598-019-39418-5. PubMed PMID: 30816188; PubMed Central PMCID: PMC6395747.

**Bissler JJ**, Zadjali F, Bridges D, Astrinidis A, Barone S, Yao Y, Redd JR, Siroky BJ, Wang Y, Finley JT, Rusiniak ME, Baumann H, Zahedi K, Gross KW, Soleimani M. Tuberous sclerosis complex exhibits a new renal cystogenic mechanism. *Physiol Rep*. 2019 Jan;7(2):e13983. doi: 10.14814/phy2.13983. PubMed PMID: 30675765; PubMed Central PMCID: PMC6344348.

**Bissler JJ**, Christopher Kingswood J. Renal manifestation of tuberous sclerosis complex. *Am J Med Genet C Semin Med Genet*. 2018 Sep;178(3):338-347. doi:10.1002/ajmg.c.31654. Epub 2018 Oct 11. Review. PubMed PMID: 30307110.

**Bissler JJ**, Nonomura N, Budde K, Zonnenberg BA, Fischereeder M, Voi M, Louveau AL, Herbst F, Bebin EM, Curatolo P, Zonta A, Belousova E. Angiomyolipoma rebound tumor growth after discontinuation of everolimus in patients with tuberous sclerosis complex or sporadic lymphangiomyomatosis. *PLoS One*. 2018 Sep 7;13(9):e0201005. doi: 10.1371/journal.pone.0201005. eCollection 2018. PubMed PMID: 30192751; PubMed Central PMCID: PMC6128468.

Atherton JG, Hains DS, **Bissler J**, Pendley BD, Lindner E. Generation, clearance, toxicity, and monitoring possibilities of unaccounted uremic toxins for improved dialysis prescriptions. *Am J Physiol Renal Physiol*. 2018 Oct 1;315(4):F890-F902. doi: 10.1152/ajprenal.00106.2017. Epub 2018 Mar 14. PubMed PMID: 29537310.

### **Brown, Chester**

Bu Y, Okunishi K, Yogosawa S, Mizuno K, Irudayam MJ, **Brown CW**, Izumi T. Insulin Regulates Lipolysis and Fat Mass by Upregulating Growth/Differentiation Factor 3 in Adipose Tissue Macrophages. *Diabetes*. 2018 Sep;67(9):1761-1772. doi: 10.2337/db17-1201. Epub 2018 Jun 26. PubMed PMID: 29945891.

### **Bukiya, Anna**

North K, Bisen S, Dopico AM, **Bukiya AN**. Tyrosine 450 in the Voltage- and Calcium-Gated Potassium Channel of Large Conductance Channel Pore-Forming (slo1) Subunit Mediates Cholesterol Protection against Alcohol-Induced Constriction of Cerebral Arteries. *J Pharmacol Exp Ther*. 2018 Nov;367(2):234-244. doi:10.1124/jpet.118.250514. Epub 2018 Aug 16. PubMed PMID: 30115756; PubMed Central PMCID: PMC6170972.

### **Carson, James**

Fix DK, VanderVeen BN, Counts BR, **Carson JA**. Regulation of Skeletal Muscle DRP-1 and FIS-1 Protein Expression by IL-6 Signaling. *Oxid Med Cell Longev*. 2019 Feb 21;2019:8908457. doi 10.1155/2019/8908457. eCollection 2019. PubMed PMID: 30918582; PubMed Central PMCID: PMC6408992.

VanderVeen BN, Fix DK, Montalvo RN, Counts BR, Smuder AJ, Murphy EA, Koh HJ, **Carson JA**. The regulation of skeletal muscle fatigability and mitochondrial function by chronically elevated interleukin-6. *Exp Physiol*. 2019 Mar;104(3):385-

397. doi: 10.1113/EP087429. Epub 2019 Jan 15. PubMed PMID: 30576589.

### **Chauhan, Subhash**

Massey AE, Sikander M, Chauhan N, Kumari S, Setua S, Shetty AB, Mandil H, Kashyap VK, Khan S, Jaggi M, Yallapu MM, Hafeez BB, **Chauhan SC**. Next-generation paclitaxel-nanoparticle formulation for pancreatic cancer treatment. *Nanomedicine*. 2019 Aug;20:102027. doi: 10.1016/j.nano.2019.102027. Epub 2019 Jun 4. PubMed PMID: 31170509; PubMed Central PMCID: PMC6705422.

2: Khan S, Setua S, Kumari S, Dan N, Massey A, Hafeez BB, Yallapu MM, Stiles ZE, Alabkaa A, Yue J, Ganju A, Behrman S, Jaggi M, Chauhan SC. Superparamagnetic iron oxide nanoparticles of curcumin enhance gemcitabine therapeutic response in pancreatic cancer. *Biomaterials*. 2019 Jul;208:83-97. doi:10.1016/j.biomaterials.2019.04.005. Epub 2019 Apr 8. PubMed PMID: 30999154.

Sikander M, Malik S, Chauhan N, Khan P, Kumari S, Kashyap VK, Khan S, Ganju A, Halaweish FT, Yallapu MM, Jaggi M, **Chauhan SC**. Cucurbitacin D Reprograms Glucose Metabolic Network in Prostate Cancer. *Cancers (Basel)*. 2019 Mar 14;11(3). pii: E364. doi: 10.3390/cancers11030364. PubMed PMID: 30875788; PubMed Central PMCID: PMC6469021.

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Hatami E, Mu Y, Shields DN, **Chauhan SC**, Kumar S, Cory TJ, Yallapu MM. Mannose-decorated hybrid nanoparticles for enhanced macrophage targeting. *Biochem Biophys Rep*. 2019 Jan 25;17:197-207. doi: 10.1016/j.bbrep.2019.01.007.eCollection 2019 Mar. PubMed PMID: 30723809; PubMed Central PMCID: PMC6351286.

Kashyap VK, Wang Q, Setua S, Nagesh PKB, Chauhan N, Kumari S, Chowdhury P, Miller DD, Yallapu MM, Li W, Jaggi M, Hafeez BB, **Chauhan SC**. Therapeutic efficacy of a novel  $\beta$ III/ $\beta$ IV-tubulin inhibitor (VERU-111) in pancreatic cancer. *J Exp Clin Cancer Res*. 2019 Jan 23;38(1):29. doi: 10.1186/s13046-018-1009-7. PubMed PMID: 30674344; PubMed Central PMCID: PMC6343279.

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Nagesh PKB, Chowdhury P, Hatami E, Boya VKN, Kashyap VK, Khan S, Hafeez BB, **Chauhan SC**, Jaggi M, Yallapu MM. miRNA-205 Nanoformulation Sensitizes Prostate Cancer Cells to Chemotherapy. *Cancers (Basel).* 2018 Aug 25;10(9). pii: E289. doi:10.3390/cancers10090289. PubMed PMID: 30149628; PubMed Central PMCID: PMC6162422.

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Chowdhury P, Nagesh PKB, Khan S, Hafeez BB, **Chauhan SC**, Jaggi M, Yallapu MM. Development of polyvinylpyrrolidone/paclitaxel self-assemblies for breast cancer. *Acta Pharm Sin B.* 2018 Jul;8(4):602-614. doi: 10.1016/j.apsb.2017.10.004. Epub 2017 Dec 10. PubMed PMID: 30109184; PubMed Central PMCID: PMC6090082.

Tripathi MK, Doxtater K, Keramatnia F, Zacheaus C, Yallapu MM, Jaggi M, **Chauhan SC**. Role of lncRNAs in ovarian cancer: defining new biomarkers for therapeutic purposes. *Drug Discov Today.* 2018 Sep;23(9):1635-1643. doi:10.1016/j.drudis.2018.04.010. Epub 2018 Apr 23. Review. PubMed PMID: 29698834; PubMed Central PMCID: PMC6139057.



## **Chen, Hao**

Zhao Q, Yang J, **Chen H**, Li J, Que L, Zhu G, Liu L, Ha T, Chen Q, Li C, Xu Y, Li Y. Peli1 induction impairs cardiac microvascular endothelium through Hsp90 dissociation from IRE1 $\alpha$ . *Biochim Biophys Acta Mol Basis Dis*. 2019 Oct 1;1865(10):2606-2617. doi: 10.1016/j.bbadis.2019.06.017. Epub 2019 Jun 29. PubMedPMID: 31260751.

DiCarlo GE, Aguilar JI, Matthies HJ, Harrison FE, Bundschuh KE, West A, Hashemi P, Herborg F, Rickhag M, **Chen H**, Gether U, Wallace MT, Galli A. Autism-linked dopamine transporter mutation alters striatal dopamine neurotransmission and dopamine-dependent behaviors. *J Clin Invest*. 2019 May 16;129(8):3407-3419. doi: 10.1172/JCI127411. PubMed PMID: 31094705; PubMed Central PMCID: PMC6668686.

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APPENDICES

Schedule 7

Schedule 7

CENTERS OF EXCELLENCE ACTUAL, PROPOSED, AND REQUESTED BUDGET

	FY 2018-19 Actual			FY 2019-20 Proposed			FY 2020-21 Requested		
	Matching	Appropri.	Total	Matching	Appropri.	Total	Matching	Appropri.	Total
<b>Expenditures</b>									
<b>Salaries</b>									
Faculty	\$0	\$0	\$0	\$0	\$0	\$0	\$0	\$0	\$0
Other Professional	76,670	152,061	228,731	48,668	155,102	203,770	51,101	160,000	211,101
Clerical/ Supporting	65,827	242,940	308,767	52,985	247,799	300,784	55,635	250,000	305,635
Assistantships	0	0	0	0	0	0	0	0	0
<b>Total Salaries (Excluding Longevity)</b>	<b>\$142,497</b>	<b>\$395,001</b>	<b>\$537,498</b>	<b>\$101,653</b>	<b>\$402,901</b>	<b>\$504,554</b>	<b>\$106,736</b>	<b>\$410,000</b>	<b>\$516,736</b>
Longevity (Exclude from Salaries)	\$1,350	\$11,337	\$12,687	\$1,418	\$10,000	\$11,418	\$1,488	\$10,000	\$11,488
Fringe Benefits	21,006	126,616	147,622	22,056	130,148	152,204	23,159	135,000	158,159
<b>Total Personnel</b>	<b>\$164,853</b>	<b>\$532,954</b>	<b>\$697,807</b>	<b>\$125,127</b>	<b>\$543,049</b>	<b>\$668,176</b>	<b>\$131,383</b>	<b>\$555,000</b>	<b>\$686,383</b>
<b>Non-Personnel</b>									
Travel	\$0	\$2,414	\$2,414	\$0	\$3,000	\$3,000	\$0	\$3,100	\$3,100
Software	0	0	0	0	0	0	0	0	0
Books & Journals	0	0	0	0	0	0	0	0	0
Other Supplies	820,000	149,237	969,237	861,000	13,045	874,045	904,050	14,985	919,035
Equipment	313,000	0	313,000	328,650	0	328,650	345,083	0	345,083
Maintenance	0	41,129	41,129	0	40,000	40,000	0	0	0
Scholarships	0	0	0	0	0	0	0	0	0
Consultants	0	0	0	0	0	0	0	0	0
Renovation	0	0	0	0	0	0	0	0	0
Other (Specify):									
Media Processing	0	1,136	1,136	0	1,100	1,100	0	1,000	1,000
Communication	0	4,825	4,825	0	5,000	5,000	0	5,000	5,000
Professional Services & Memberships	30,000	110,276	140,276	31,500	90,000	121,500	33,075	94,000	127,075
Rentals & Insurance	0	360	360	0	370	370	0	350	350
Contractual & Special Services	0	-232,674	-232,674	0	0	0	0	0	0
<b>Total Non-Personnel</b>	<b>\$1,163,000</b>	<b>\$76,703</b>	<b>\$1,239,703</b>	<b>\$1,221,150</b>	<b>\$152,515</b>	<b>\$1,373,665</b>	<b>\$1,282,208</b>	<b>\$118,435</b>	<b>\$1,400,643</b>
<b>GRAND TOTAL</b>	<b>\$1,327,853</b>	<b>\$609,657</b>	<b>\$1,937,510</b>	<b>\$1,346,277</b>	<b>\$695,564</b>	<b>\$2,041,841</b>	<b>\$1,413,591</b>	<b>\$673,435</b>	<b>\$2,087,026</b>
<b>Revenue</b>									
New State Appropriation	\$0	\$632,516	\$632,516	\$0	\$641,367	\$641,367	\$0	\$673,435	\$673,435
Carryover State Appropriation	0	31,338	31,338	0	54,197	54,197	0	0	0
New Matching Funds	1,327,853	0	1,327,853	1,346,277	0	1,346,277	1,413,591	0	1,413,591
Carryover from Previous Matching Funds	0	0	0	0	0	0	0	0	0
<b>Total Revenue</b>	<b>\$1,327,853</b>	<b>\$663,854</b>	<b>\$1,991,707</b>	<b>\$1,346,277</b>	<b>\$695,564</b>	<b>\$2,041,841</b>	<b>\$1,413,591</b>	<b>\$673,435</b>	<b>\$2,087,026</b>

Institution: UNIVERSITY OF TENNESSEE HEALTH SCIENCE CENTER Center: MOLECULAR RESOURCE CENTER

**Molecular Resource Center of Excellence Personnel,  
FY19 (2018-2019)**

Institution:	The University of Tennessee Health Science Center, Memphis, TN
Center:	Molecular Resource Center
Tiffany Seagroves, Ph.D.	Executive Director
William Taylor, Ph.D.	Director; Microarray, Whole Genome Sequencing and RNA analysis
Tom Cunningham, Ph.D.	Associate Director; DNA sequencing, genotyping, general molecular biology
Felicia Waller	Specialist; Fluorescence microscopy, sample quality control, DNA sequencing, RT PCR, assist with invoicing customers
Lorne Rose	Senior Research Specialist; Microarrays technician/analyst and NGS assistance
Vacant	Senior Research Assistant; Whole Genome sequencing (position vacated April 2017 and reclassified December 2017)
Natalie Smith	Business Manager (January 2017-present)

**Personnel Summary**

Appropriated costs for 2018-2019	\$532,954
Matching funds for 2018-2019	\$164,853
<b>Total costs for 2018-2019</b>	<b>\$697,807</b>

**Molecular Resource Center of Excellence  
Proposed Core Personnel, FY20 (2019-2020)**

Institution:	The University of Tennessee Health Science Center, Memphis, TN
Center:	Molecular Resource Center
Tiffany Seagroves, Ph.D.	Executive Director and Associate Vice Chancellor for Research-Core Labs
William Taylor, Ph.D.	Director; Microarray, Whole Genome Sequencing and RNA analysis
Tom Cunningham, Ph.D.	Associate Director; DNA sequencing, genotyping, general molecular biology
Vacant	Senior Research Assistant, Microarrays
Lorne Rose	Senior Research Specialist; Microarrays technician/analyst and NGS assistance
Felicia Waller	Specialist; Fluorescence microscopy, sample quality control, DNA sequencing, RT-PCR, assist with billing
Natalie Smith	Business Manager

**Personnel Summary**

Appropriated funds for 2019-2020	\$543,049
Matching funds for 2019-2020	\$125,127
<b>Total funds for 2019-2020</b>	<b>\$668,176</b>

**Molecular Resource Center of Excellence  
Non-Personnel Actual and Proposed Budget Summaries  
and Actual Appropriated Funds and Matching Funds Summary**

**Actual 2018-2019 Non-Personnel Total Budget**

Travel	\$ 2,414
Software	\$ 0
Supplies	\$ 969,237
Equipment	\$ 313,000
Maintenance	\$ 41,129
Contract/Special Services	\$ (232,674)
Prof Services and Membership	\$ 140,276
Communications	\$ 4,825
Media	\$ 1,136
Rentals	<u>\$ 360</u>
<b>Total</b>	<b>\$1,239,703</b>

**Non-Personnel Summary**

Appropriated funds for 2018-2019	\$ 76,703
Matching funds for 2018-2019	\$1,163,000
<b>Total funds for 2018-2019</b>	<b>\$1,239,703</b>

**Proposed 2019-2020 FY20 Non-Personnel Budget**

Travel	\$ 3,000
Software	\$ 0
Supplies	\$ 874,045
Equipment	\$ 328,650
Maintenance	\$ 40,000
Contract/Special Service	\$ .....0
Prof Services and Membership	\$ 121,500
Communication	\$ 5,000
Media	\$ 1,100
Rentals	\$ 370
Other	<u>\$ 0</u>
<b>Total</b>	<b>\$1,373,665</b>

**Non-Personnel Summary, Proposed**

Appropriated funds for 2019-2020	\$ 152,515
Matching funds for 2019-2020	<u>\$1,221,150</u>
<b>Total funds for 2018-2019</b>	<b>\$1,373,665</b>



**Actual 2018-2019 Appropriated Funds and Matching Funds**

Total Appropriated Funds 2018-2019	\$ 632,516
Carry Over of 2017-2018 funds	\$ 31,338
Total Appropriation 2018-2019	\$ 663,854
Total Matching funds 2018-2019	<u>\$1,327,853</u>
<b>Total Operating Budget, 2018-2019</b>	<b>\$1,991,707</b>